LOI510, containing *PET* genes and *cat* within the *E. coli pfl* gene was digested with *Sal1*, or *Pst1*, circularized and transformed into *K. oxytoca* strain M501. See Wood *et al.*, *Appl. Environ. Microbiol.* 58:2103 (1992), which is herein incorporated by reference. Additionally, homologous *K. oxytoca* M5A1 DNA prepared by *Sau3*A digestion was ligated to a 4.6 kb *Bam*H1 fragment containing only the *PET* and *cat* genes was also used for transformation. Recombinants were initially selected using 20 μg chloramphenicol/ml and expressed low levels of *Z. mobilis* enzymes. As with *E. coli* (Ohta *et al.* [1991] *Appl. Environ. Microbiol.* 57: 893-900), expression was boosted by direct selection of mutants with resistance to 600 μg chloramphenicol/ml. A single clone expressing high level resistance was retained for each independent integration event.

In the Claims:

Please cancel without prejudice or disclaimer claims 6, 17-24, 25-34, 40, 42-45 and 47.

Please amend claims 48 and 49 as follows:

48. (Amended) The recombinant host cell strain according to claim 1, of *Klebsiella oxytoca* M5A1comprising a plasmid represented by a deposit with the American Type Culture Collection designated as deposit number ATCC 68564.

49. (Amended) A cell strain according to claim 11, wherein said cell strain is designated P2 and represented by a deposit with the American Type Culture Collection designated as deposit number ATCC 55307.